

# Flavio Di Pisa, PhD

**Institute of Biophysics, National Research Council,  
Via Corti, 12 - 20133 Milan**

## Professional Profile

Structural biologist with a decade of comprehensive experience in the field. Proficient in structure-function studies of macromolecular complexes. Adept in employing cutting-edge techniques and methodologies to unravel structural puzzles.

## Personal Information

## Professional career

### 2022-Ongoing

#### Principal Investigator

Institute of Biophysics, National Research Council (CNR), Milan  
Lab Head: Dr. Eloise Mastrangelo

### 2020-2022

#### Post-Doctoral Fellow

Biocrystallography unit, San Raffaele Research Hospital, Milan  
Group Leader: Prof. Massimo Degano

### 2017-2020

#### Post-Doctoral Fellow

Structural Vaccinology Lab, University of Milan  
Group Leader: Prof. Martino Bolognesi  
Lab Head: Prof. Louise Gourlay

### 2015-2017

#### Post-Doctoral Fellow

Protein Crystallography and Structural Biology Lab, University of Siena  
Group Leader: Prof. Stefano Mangani

## Experience as visiting Researcher

**University of Halle** (october 2024 – december 2024) : Cryo-EM data acquisition and processing, Prof. Kastitis Laboratory ([link](#) , section Visiting Professors/Researchers)

**ESRF synchrotron, Grenoble** (7/02/2025 – 9/02/2025, 15/12/2017 – 16/12/2017, 11/02/2018 – 13/02/2018, 25/07/2018 – 27/07/2018, 16/11/2016 – 18/11/2016, 13/04/2016 – 16/04/2016, 27/09/2012 – 29/09/2012, 31/05/2012 – 02/06/2012) : data acquisition using SAXS and single crystal diffraction techniques

**Institut Laue-Langevin, Grenoble** (22/05/2024 – 24/05/2024): data acquisition using SANS technique,

**DLS, Diamond Light Source, Oxfordshire** (22/01/2017 – 24/02/2017, 08/07/2016 – 10/07/2016, 28/07/2015 – 30/07/2015, 03/05/2015 – 05/05/2015, 17/02/2015 – 19/02/2015, 02/05/2014 – 04/05/2014, 02/02/2014 – 04/02/2014, 15/05/2013 – 17/05/2013, 04/07/2013 – 06/07/2013) : data acquisition using single crystal diffraction techniques

## Teaching work experience

### 2021- 2022

Teacher of Medical Chemistry, Dental School - Master's Degree in Dentistry and Dental Implantology, San Raffaele Research Hospital, Milan

### 2017-2018

Teaching assistant, University of Milan  
Wet lab tutoring for internship in Experimental Biology, Referent: Prof. Louise Gourlay

### 2018-2019

Teaching assistant, University of Milan  
Wet lab tutoring for internship in Experimental Biology, Referent: Prof. Louise Gourlay/ Prof. Monica Beltrame

## Education and training

**2015:** PhD in Chemical and Pharmaceutical Sciences, University of Siena (Italy)  
Department of Biotechnology, Chemistry and Pharmacy  
Supervisor: Prof. Stefano Mangani

*Thesis entitled: "Crystallographic studies to unravel reaction mechanisms and enzyme-substrate interactions in target proteins"*

**2011:** Master Degree in Chemistry, awarded with 110/110 marks cum laude, University of Siena (Italy)  
Supervisor: Prof. Stefano Mangani

*Thesis entitled: "Structure-function studies through Crystallographic Methods of class-D  $\beta$ -Lactamases"*

**2009:** Bachelor Degree in Chemistry, awarded with 106/110 marks, University of Siena (Italy)  
Supervisor: Prof. Stefano Mangani

*Thesis entitled: "High resolution refinement of the three- dimensional structure of the TRU-1 enzyme from Aeromonas Enteropelogenes".*

## Funding for research

### IRONYc: IRON homeostasis in Neuroferritinopathy: unravelling the molecular details of abnormal ferritin functionality

Project funded by **Fondazione Cariplo**, 250 kEU (2022-2026)

#### Coordinator: Dr. Flavio Di Pisa (CNR, Milano, Italy)

Partners: Prof. Sonia Levi (UniSR, Milano, Italy) and Prof. Maria Luisa Gelmi (Unimi, Milano, Italy)

## Languages

- Italian: mother tongue.
- English: spoken (fluent) and written (advanced).

## Expertise and competences

**Molecular Biology and Microbiology:** vector design and high- throughput cloning techniques (Overlap Extension PCR, SLIC), isolation and transformation, DNA purification (mini- and maxi- prep), DNA gel analysis, SDS-PAGE, western blotting,

**Protein Expression and Purification:** bacterial protein expression (*Escherichia Coli*), eukaryotic protein expression (baculovirus- infected insect cells), AKTA HPLC Systems (Pure, Explorer, Start), chromatographic methods (IMAC/SEC/IE), affinity tag purification (MBP, SUMO, Chitin Binding Domain).

**Protein characterization & Biophysics:** MALDI-TOF MS, UV-Vis spectroscopy, Circular Dichroism (CD), Fluorescence Spectroscopy, dynamic light scattering DLS, Differential Scanning Fluorimetry (DSF)

**Crystallization techniques:** Vapor diffusion (hanging drop, sitting drop), microbatch under oil and optimisation of crystallisation conditions. Use of automatic pipetting robots Oryx 4 and Oryx 8 (Douglas Instruments).

#### Protein X-ray Crystallography:

- X-ray data collection at synchrotrons (ESRF, DLS, Elettra)
- Data processing (XDS, iMosflm)
- Structure determination by Molecular Replacement
- Model Building (Autobuild, Buccaneer) and structure refinement (Refmac, Buster, Phenix refine)
- Structure validation (Molprobtity, PDB Redo)
- Extensive knowledge of technical softwares for X-ray structure determination (CCP4 suite, Phenix, SHELX) and visualization (Pymol, CCP4mg, Chimera).

#### Cryo Electron Microscopy:

- Single particle cryo-electron microscopy data processing (CryoSPARC software)
- Model building using real space refinement (Phenix package)

#### Computer proficiency:

- Office Suite (word processor, spread sheet, presentation software) and Internet search engines (Chrome, Explorer, Firefox).
- Basic knowledge of Bash programming language (Linux).